The genus *Anthelephila* (Coleoptera: Anthicidae)

**Key words.** Taxonomy, redescription, new synonymy, lectotype designation, Coleoptera, Anthicidae, *Anthelephila*

**Abstract.** The genus *Anthelephila* Hope, 1833 and its type species, *Anthelephila cyanea* Hope, 1833, are redescribed. Based on examination of the type material, the following new synonymy is proposed, *Anthelephila* Hope, 1833 (= *Formicoma* Motschoulsky, 1845 syn. n.) and *A. cyanea* Hope, 1833 (= *Notocux caeruleus* Thunberg, 1787 syn. n.). Five genus-group names are regarded as unjustified emendations and are placed as synonyms: *Anthelephila* Hope, 1833 (= *Anthelephilus* LaFerté-Sénectère, 1849; *Formicosa* Motschoulsky, 1845; *Myrmecosoma* Mannerheim, 1846; *Formicomus* LaFerté-Sénectère, 1849; *Orthauchen* Krekich-Strassoldo, 1925 syn. n.). A lectotype is designated for *Anthelephila cyanea* Hope, 1833.

**INTRODUCTION**

There are two closely related genera currently placed in the Formicomini: *Anthelephilus* LaFerté-Sénectère, 1849, comprising about 20 species, and *Formicomus* LaFerté-Sénectère, 1849 comprising more than 350 species. Unfortunately, the present nomenclatorial status of both these names is in serious contradiction with the ICZN (1999) rules, since they were knowingly and unjustifiedly proposed by Hope (1840) and LaFerté-Sénectère (1849a, b) to replace the older names *Anthelephila* Hope, 1833 and *Formicoma* Motschoulsky, 1845, with new spellings for these two genera accepted by nearly all subsequent authors.

The present paper is aimed to solve the above mentioned problem, and to provide a redescription of the genus *Anthelephila* Hope and its type species. Furthermore, brief remarks are made upon the general topics of biology, geographic distribution, and position of *Anthelephila* within the subfamily Anthicinae.

**MATERIAL AND METHODS**

Specimens were examined with a stereoscopic microscope using diffused lighting, and illustrations were made using an ocular microgrid. Morphological measurements were taken using the ocular micrometer. Male and female genitalia were illustrated after having been cleared by boiling in 10% KOH solution.

Author’s comments on the type material are found in square brackets. Exact label data are quoted for the type specimens only. Separate labels are indicated by slashes (\). The publication date of the papers by LaFerté-Sénectère follows Chandler (2000).

**DEPOSITORIES**


MNHB - Museum für Naturkunde der Humboldt-Universität, Berlin, Germany.


UZIU - Uppsala University, Zoological Museum, Uppsala, Sweden.

CKD - collection Zbyněk Kejval, Domažlice, Czech Republic.

CUP - collection Gerhard Uhmann, Pressath, Germany.

DSC - collection Donald S. Chandler, Durham, New Hampshire, U. S. A.

**TAXONOMY**

**TAXONOMIC HISTORY OF THE TREATED GENERIC NAMES**

*Anthelephila*, *Anthelephilus*. The genus *Anthelephila* was originally established by F. W. Hope as a subgenus of *Anthicus* Paykull, 1798. The description was initially read at the meeting of the Zoological Society in London on May 28th 1833, and then published as a report of this meeting in an abbreviated format (comprising only more important characters of the genera and species described) in July of the same year in the *Proceedings of Zoological Society of London* (Hope, 1833). The full description was published the next year (Hope, 1834) in the *Transactions of the Zoological Society of London*. Since the description in the *Proceedings* is clearly attributed to Hope and lists several characters for recognition of the genus *Anthelephila* and its type species, *Anthicus cyaneus* Hope, it is regarded as the original description (cf. Sherborn, 1928; Neave, 1939). Saunders (1834) added another two species and was the first to treat *Anthelephila* as a genus, without providing any comments on his rationale for this elevation. A change in gender to *Anthelephilus* was first used, without comments, by Hope (1840) to replace *Anthelephila*. Although he used *Anthelephilus* in combination with Hope’s authorship, LaFerté-Sénectère (1849a, c) was apparently quite unaware of the preceding paper by Hope, and proposed more formally the same emendation, along with a generic description.

According to the ICZN (1999), *Anthelephilus* is regarded as unjustified emendation of *Anthelephila*. After 1899, *Anthelephilus* was used as valid name by Krekich-Strassoldo (1931), Sakai (1989) and Lafer (1996).

**Formicoma, Formicosoma, Formicomus.** The genus *Formicoma* was established without description by Motschoulsky (1845a) to accommodate three Palaearctic species of *Anthicus* Paykull, 1798. Subsequently, Motschoulsky (1845b) emended *Formicoma* to *Formicosoma* in a list of corrections to the preceding paper.

The name *Formicomus* was established by LaFerté-Sénectère to replace *Formicoma*. This fact is evident from the following statement: “...depuis, ayant appris que M.V. Motschoulsky avait également séparé ces insectes des Anthicus dans le catalogue de sa collection actuellement sous presse, et qu’il leur avait donné le nom Formicomus, nous avons cru devoir, pour simplifier la nomenclature, adopter le nom de M. Motschoulsky, en lui donnant une terminaison masculine” (footnote in LaFerté-Sénectère, 1849b: 2).

According to the ICZN (1999), both *Formicosoma* and *Formicomus* are unjustified emendations of *Formicoma*. On the other hand, *Formicoma* was used later as a valid name only by Kolenati (1846) and Motschoulsky (1849), and the youngest name *Formicomus* is undoubtedly in prevailing usage and could thus be conserved (ICZN, 1999: Article 23.9.1.1–2). This action, however, would be unnecessary for the reasons described below.

**Myrmecosoma.** The name *Myrmecosoma* was proposed by Mannheim (1846) to replace *Formicoma*, since he believed the latter name to be incorrectly composed. It was used later only by Truqui (1855) and usually has been treated as synonym of *Formicoma*. According to the ICZN (1999), it is regarded as unjustified emendation of *Formicoma*.

**Orthauchen.** The subgenus *Orthauchen* was established by Krekich-Strassoldo (1925) to accomodate the species of *Formicomus* with long neck smoothly merging with head. It was recently synonymized with the nominotypical subgenus by Kejval (2000).

**Carteromerus.** Pic (1911) listed *Carteromerus* as synonym of *Formicomus* with the following data: “*Carteromerus* Laf. (Monogr. 1848, p. 71, nota 1).” According to the respective note (pages 2, 71) in LaFerté-Sénectère (1849b, c), *Carteromerus* is the original name LaFerté-Sénectère meant to use before he decided to emend Motschoulsky's *Formicoma*. It occurs on the original labels in the LaFerté-Sénectère’s collection, however it was never officially established since there were no explicitly included species, and he never used it as a valid name. Consequently, it is herein regarded as an unavailable name.

**Comments on the newly proposed synonymy**

The only character traditionally used to separate the genera *Formicomus* and *Anthelephilus* is the absence of hind membranous wings in the latter genus, which is related to the obsolete elytral humeri, and sometimes also to the truncate elytral apices. This difference was first noted by LaFerté-Sénectère; although he had not seen the type specimen of *Anthelephilus*, he assumed its aptery solely from a copy (Fig. 8) of the original habitus illustration (Fig. 9) provided by Chevrolat (LaFerté-Sénectère, 1849a: 1). However, as was already stressed by some early workers (Lacordaire, 1859; King, 1869), there is no supporting evidence for this division. The reduction of hind wings is well known in numerous genera of Anthicidae, and their tendency toward reduction has been observed in a group of closely related species of *Formiconus* (Kejval, 2000). The most widely known species treated under the name *Anthelephilus* (A. ruficollis Saunders, 1834) appears to vary in this character as I have seen fully winged, brachypterous, and apterous specimens. Finally, examination of the type material has shown that the type species of the genus *Anthelephilus* is not apterous (see the redescription of *A. cyaneus* below).

Motschoulsky (1845a) designated “*Anth. pedestris* F.” as the type species of *Formicoma*. Why Motschoulsky and Dejean (1837) attributed this species to Fabricius is not clear. It is evident from the data given by Fabricius (1801), that he just proposed one new combination for the species described as *Carabus pedestriv* by Rossi (1790), and accordingly *Anthus pedestriv* is not listed as a Fabrician species by Zimens (1964). The major part of the collection of P. Rossi is now deposited in MNHB as a part of the so called “Historical collection,” and present there is the series labelled “Pedestriv R.* [R. == Rossi, * = type material], Anthicus. pedestriv Fab., N. thoracicus Pz.,” comprising specimens of various origin, including the type(s) of *Carabus pedestriv* Rossi, 1790. Having examined the whole series, I found it homogeneous and belonging to *Formicomas pedestriv* (Rossi, 1790) in its present sense, which is a well known species clearly congenic with *Anthelephilus cyaneus*. Considering these facts, *Formicoma, Formicosoma, Myrmecosoma* and *Formicomus* are all regarded as junior synonyms of *Anthelephilus*.

**Genus Anthelephilus Hope, 1833**


**Type species.** *Anthicus (Anthelephilus) cyaneus* Hope, 1833; fixed by monotypy.

*Anthelephilus* LaFerté-Sénectère, 1849a: 1, syn. n.

*Antellephilus*; Pic, 1901: 249. Subsequent misspelling of *Anthelephilus*.

*Antellephilus*; Pic, 1899: 758. Subsequent misspelling of *Anthelephilus*.
Anthelephila; Krekich-Strassoldo, 1929: 475. Subsequent misspelling of *Anthelephus*.

*Formicosa* Motschoulsky, 1845a: 83, syn. n.

**Type species.** *Carabus pedestris* Rossi, 1790 (as *Anthicus pedestris*); fixed by original designation.

*Formicosoma* Motschoulsky, 1845b: unnumbered erratum page, syn. n.

*Myrmecosoma* Mannerheim, 1846: 227, syn. n.

*Formicomus* LaFerté-Sénectère, 1849b: 1, syn. n.

*Orthothen* Krekich-Strassoldo, 1925: 516, syn. n.

**Type species.** *Formicomus aestimabilis* Krekich-Strassoldo, 1919; fixed by subsequent designation by Kejval (2000).

*Carteromerus* [nomen nudum].

**Diagnosis.** Frontoclypeal sulcus fine but distinct. Prothorax with distinct apical flanged rim (collar), more or less prominent lateral antebasal impression in posterior half and with basal margin distinctly bordered dorsally by a thin, well defined sulcus originating at lateral foveae. Mesosternum triangular, anterior angle of mesosternum distant from anterior margin of mesothorax due to the mediadly broadly joined mesepisterna (Figs 1, 6), lateral margins either moderately arcuate or straight, intercoxal process fully separating mesocoxae; lateral margins of mesepisterna simply connected (not raised above level of) with mesepimera (Figs 2, 11), bare, without distinct fringe of setae. Mesothoracic gland conspicuously large, differentiated into paired canal and reservoir and its opening sunk into mesothorax (Hemp, 1994). Mesocutellum longitudinally triangular, tapering posteriorly (Fig. 21). Metendosternite with narrow arms and obsolete ventro-lateral projections (lamina) (Fig. 12). Elytra in males lacking apical notches; sutural striae very vaguely indicated in apical third. Antennal insertion exposed and clearly visible, sockets sometimes slightly hidden by moderately raised lateral margin of frons in dorsal view. All femora distinctly clavate, metafemora simply shaped; all tibiae with two well developed apical spurs (most prominent on metatibiae). Abdominal sternum III without foveae immediately behind metacoxae, its intcroctal process widely, somewhat irregularly rounded apically and incompletely bordered in ventral view (Figs 3, 4); male abdominal segment VIII modified, sternite with a pair of posteriorly projecting progs; male sternite IX (spiculum) formed by comparatively fine, fibre-like apodeome, bifurcate posteriorly, Y-shaped (Fig. 28). Male aedeagus less sclerotized, tegmen clearly divided into the apical portion (formed by fused parameres) and basipiece; median lobe strengthened by simple, rod-like, sclerotized apodeme, gonopore (of rather uniform morphology) situated at level or shortly behind basal margin of tegmen (Fig. 29). Female ovipositor (Fig. 19) with well developed coxites and styli.

**Sexual dimorphism.** Adults of most species of *Anthelephila* show rather distinct sexual dimorphism. Males are usually of smaller size and exhibit a more slender body-form than females, sometimes have slightly longer antennae, and display a number of conspicuous secondary sexual characters: front legs almost exclusively modified, with variously shaped processes, protuberances (very rarely present also on meso- or metatibiae); metasternum and abdominal sterna III-VI often with longitudinal ribs, protrusions or protuberances, impressions, densely punctured and setose patches; abdominal sternum VII with posterior margin variously shaped (very rarely simple), often more or less emarginate, often with median process of various form and length; sternite VIII always modified and mostly of rather complicated morphology, its paired prongs armed with various processes, lobes, clustered setae etc.; metatibiae and metatarsi sometimes with longer, bristle-like setae.

Females are generally more simple in appearance: legs, metasternum and abdominal sterna III-VI always simple; sternum and tergum VII predominantly simply shaped, rarely somewhat prolonged and more tapering, sometimes truncate or notched apically; segment VIII weakly sclerotized, composed of fine simple sclerites, more or less conspicuously setose along posterior margin. Sometimes, females may differ from males by the modified mesosternum (see Figs 92–102 in Kejval, 2000), but in general they very rarely possess a modifications not found in their respective males, e.g. the prolonged and tapering elytral apices (few Oriental species) or the densely punctured and setose patches on the pronotum (*A. picta* Kejval, 2002).

**Immature stages.** The first instar of larva of *Anthelephila pedestris* (Rossi, 1790) was described by Schütz (1989).

**Biology.** *Anthelephila* appears to be relatively thermophilous, with the majority of species occurring in the tropics of Africa and southeast Asia, and only few species penetrating northwards into the Eurosiberian subregion. Its members inhabit both open and forested habitats at elevations ranging from sea level up to 3500 m (in the Himalaya Mts.). They are mostly collected by sweeping vegetation or beating foliage along trails, or at the margins of clearings.

Both larvae and adults of *Anthelephila* are apparently scavengers and opportunistic predators, feeding on organic debris and small or weakened invertebrates, as it is typical of most other genera of Anthicidae (Werner & Chandler, 1995). Mycophagy was observed by Hemp et al. (1997) for adults of two species in Tanzania.

At this time fourteen species of *Anthelephila* are known to be canthariphilous, that is they are known to be attracted to the terpenoid cantharidin (Hemp & Dettner, 2001). This substance serves as a mating attractant, and perhaps functions as an aggregation pheromone. In addition, as a toxic component of haemolymph, it provides protection for both adults and immature stages (Hemp et al., 1997, Hemp & Dettner, 1997). Based on field observations and chemical analyses (Hemp & Dettner, 1997), adults of *Anthelephila* are effectively protected by iridoids secreted by the mesothoracic glands.

Species of *Anthelephila* may resemble ants in appearance and pattern of movement, as have been noted in species of other groups of antelics (Werner & Chandler, 1995). Various species of *Anthelephila* (as *Formicomus*)
have been found running together with ants (Hemp & Dettner, 1997; Hingston, 1925; Wasmann, 1898), even near their nests, but only a few species may be potentially regarded as myrmecophilous (Wasmann, 1898).

**Distribution.** Considering the herein proposed synonymy, *Anthelephila* contains nearly 400 species distributed in the Palearctic, Afrotopical, Oriental and Australian regions. It is absent from the New World; all species of *Formicomus* described from Chile belong to different genera (Werner, 1966, 1974). The genus is extremely diverse in southeast Asia, but I know of only a few species that are east of Wallace’s line and occurring in Australia. All Australian species belonging to *Anthelephila* (about half of known Australian “*Formicomus*”) show a clear relationship to some of their Oriental congeners, and they are most frequent in the humid northern and northwestern regions of Australia. The species with dentate metafemora, which predominantly inhabit the southern arid regions of Australia, belong in my opinion to the genus *Chileanthicus* Werner, 1966 (Kejval, in prep.). The only species known from New Caledonia, *F. australocedonicus* Montouzier, 1854, belongs to a different, unrelated genus.

**Relationships.** Based on male characters, Bonadona (1974) established the tribe Formicomini for the genera *Formicomus*, *Anthelephilus*, *Stenidius* LaFerté-Sénectère, 1847 and *Andrahomanus* Pic, 1903, which all have sternite VIII of the males strongly modified. This tribe was both omitted (Ulmann, 1976) and accepted (Bucciarelli, 1980; Hemp, 1994) by subsequent authors. Although I am not quite convinced of the status of the tribe Formicomini, the following three characters may represent synapomorphies suggesting (in combination) the placement of *Anthelephila* with *Stenidius*, *Andrahomanus* and *Chileanthicus*: i) mesepisterna simply connected with mesepisterna, their margins not raised and bare (Fig. 11); ii) the intercoxal process of abdominal sternum III widely rounded to subtruncate and incompletely bordered (Fig. 4); iii) male sternite VIII modified, with distinct posteriorly projecting prongs.

In the other genera of Anthicinae that I have examined the posterolateral margins of mesepisterna were found to be completely (as in Fig. 20) or at least partly raised in the posterior half above the more or less recessed anterior portion of mesepisterna. Similarly, I failed to find a species from any of the other genera sharing both of the following two characters. The examples where one of these characters occurs represent, in my opinion, convergent evolution rather than evidence of relationship. As far as I know, only males of *Acanthinus* LaFerté-Sénectère, 1849 and some *Vacusus* Casey, 1895 possess forms of sternite VIII that may resemble the structures developed as in the above mentioned genera (see for example Fig. 43 by Werner 1970). However, the intercoxal process of abdominal sternum III in these two genera is always acutely angulate apically and with the margins completely bordered; representatives of all major groups of the large genus *Acanthinus* were examined, including the Australian species (D.S. Chandler, pers. comm.). On the other hand, the comparatively wide, apically rounded intercoxal process of abdominal sternum III occurs in various genera of Anthicinae and it can be even extremely finely, indistinctly bordered apically (although very rarely). However, sternite VIII in males of the respective species was always found to be formed by a simple flattened sclerite, with the posterior margin at most emarginate to incised medially. The genera *Phalantias* Heberdey, 1936 from South India and Sri Lanka, and *Falsoformicomus* Pic, 1948 from Madagascar, were tentatively included in the Formicomini by Hemp (1994), but share neither of the mentioned synapomorphies and exhibit other striking differences (see Bonadona, 1958, 1982).

Within the group of related genera, *Anthelephila* differs by the following major characters: anterior angle of mesocoxal sternum distant from anterior margin of mesosternum due to the medially broadly joined mesepisterna (angle situated at least very near this margin in the remaining three genera); all femora clavate (meso- and metafemora narrow in *Stenidius*); ovipositor with well developed styli (ovipositor without apparent styli in *Chileanthicus* and *Andrahomanus*).

**Anthelephila cyanea** Hope, 1833

(Figs 1-19, 21-30)

*Anthicus* (*Anthelephila*) *cyaneus* Hope, 1833: 63.

*Notoxus caerules* Thunberg, 1787: 45, syn. n.

**Redescription.**

**Body.** Total length. 3.2–4.1 mm (lectotype 3.9 mm).

**Colouration.** Black, head and pronotum with very slight bluish tinge, elytra with very distinct blue metallic reflection.

**Head.** Head 1.2 times as long as wide, moderately widely rounded posteriorly in dorsal view; tempora acutely narrowing towards base; hind temporal angles rounded; base distinctly differentiated from short neck; dorsal face of head evenly vaulted; frontal region slightly, longitudinally impressed on each side along slightly raised lateral margins. Dorsal surface distinctly, largely simply punctured; interspaces among punctures much wider than their diameter, without traces of corrugation, mostly smooth, finely punctured postero-laterally; pubescence rather short, composed of decumbent setae and few, sparsely scattered, suberect to erect setae.

Antennae moderately long, distinctly exceeding elytral humeri, distinctly enlarged and flattened distally; antennomeres I thick, 1.7 times as long as wide; antennomere II shortest, 1.2 times as long as wide; antennomeres III-VII each about twice as long as wide; antennomere X 1.2 times as long as wide; antennomere XI twice as long as wide, slightly narrowed at about midlength and bluntly pointed apically. Antennomeres mostly sparsely setose, with several longer erect setae towards apices; distal antennomeres VIII-XI much more densely and shortly setose, with erect setae less conspicuous.

**Thorax.** Pronotum 1.5 times as long as wide, as long as and distinctly narrower than head including eyes, widest before midlength, distinctly constricted (impressed laterally) in posterior half; anterior portion somewhat angularly shaped (Figs 5, 6), with rounded edge and distinct incision laterally to collar, surface behind collar flattened to slightly concave dorsally and dorso-laterally. Dorsal
f parchment of pronotum similar to that of head, slightly denser mid-dorsally; antero-lateral sides with only few, very fine punctures dorso-laterally, largely smooth and glossy, bottom of latero-basal impressions shortly, finely wrinkled and small vaulted area situated posterior to impressions finely punctured.

Mesosternum (Figs 1, 2) modified, equal in both sexes; distinctly impressed along antero-lateral margins and with rather distinct, rounded postero-median bulge situated at level of anterior margins of mesocoxae; paired antero-lateral impressions shallowly interconnected medially and becoming gradually more prominent laterad; surface largely smooth and glossy, only the intercoxal process more distinctly punctured and setose. Metasternum simple in both sexes, only shallowly impressed postero-medially.

Elytra 1.5 times as long as wide, less elongate, rather convex, moderately transversely impressed dorsally in post-scutellar area in basal third, somewhat subtruncate apically; humeri less protruding but distinct. Surface rather glossy; punctuation double, setigerous punctures sparser (especially in basal third) than on dorsal side of head and pronotum, and finer, except for few coarse punctures scattered in post-scutellar area and bearing longer tactile setae, interspaces among setigerous punctures very finely sparsely punctured, asetose; pubescence rather short, at most slightly longer than on head and pronotum, becoming denser and shorter towards elytral apices, composed of subdecumbent to nearly appressed short setae and slightly longer erect setae, with some distinctly longer suberect to erect setae in post-scutellar area. Membranous wings well developed (Fig. 16).

Legs. Penultimate tarsomere of all tarsi flattened distally, with apical margin slightly emarginate and with terminal tarsomere articulated on its dorsal side, distinctly before apex. Male front legs modified (Fig. 22); pro-
Figs 5–10. Anthelephila cyanea (female). 5 – habitus, dorsal view; 6 – thorax with head, ventral view; 7 – prothorax, lateral view; 8 – habitus (after LaFerté-Sénectère, 1849c); 9 – habitus (after Hope, 1833); 10 – the original labels of the Lectotype. Scale bars (0.5 mm): A – Figs 6, 7; B – Fig. 5.

Male abdomen. Sternum VII (Fig. 23) with posterior margin shallowly emarginate medially and with short and wide, apically rounded process projecting from dorsal side of sternum closely before posterior margin; margins of median process bearing numerous long and stiff setae. Tergum VII (Fig. 24) simply and somewhat widely rounded posteriorly, with a pair of transverse, densely spinulose patches near base. Sternite VIII (Figs 25–27)

femora moderately dilated on inner side proximally; protibiae with small, blunt, dent-like protrusion in distal half. Femora finely and comparatively sparsely setose, setae appressed to subdecumbent; tibiae (especially pro- and mesotibiae) distinctly more densely setose than femora; terminal margins of tibiae (especially metatibiae) with fringe of stiff, pointed, black setae laterally; tarsi with numerous longer, black bristle-like setae.
Figs 11–20. 11–19. Anthelephila cyanea (specimen from Greyton). 11 – mesothorax, ventro-lateral view; 12 – metendosternite; 13 – mandible, ventral view; 14 – maxilla; 15 – labium; 16 – membranous wing; 17 – female abdominal segment VIII, ventral view; 18 – the same, dorsal view; 19 – female ovipositor. 20 – Stricticomus transversalis (Villa & Villa, 1833), mesothorax, ventro-lateral view. Scale bars (0.5 mm): A – Fig. 12; B – Fig. 16. (0.1 mm): C – Fig. 15; D – Fig. 19; E – Figs 11, 13, 14, 17, 18, 20.

clearly differentiated into five sclerites: paired prongs slim, moderately divergent distally, with three longitudinal, rather densely setose edges on medial and ventral side, each prong armed apically with three more sclerotized and somewhat pointed projections and with two membranous lobes; median sclerite rather wide, transverse, with posterior margin straight, slightly emarginate medially; paired latero-basal sclerites medium-sized,
moderately dilated on medial side. Tergite VIII (Fig. 28) composed of a pair of flattened sclerites narrowly connected medially, simply rounded and long setose posteriorly.
Aedeagus (Fig. 29); tegmen with apical portion 0.7 times as long as basal-piece, rather distinctly trilobed apically, lateral lobes divergent, rounded and slightly enlarged apically, middle lobe bluntly pointed and moderately shorter than lateral lobes; median lobe of aedeagus terminating in a pair of small, membranous projections.

**Female abdomen.** Sternum VII (Fig. 30) simply shaped, with posterior margin at most slightly produced medially; tergum VII similar to that in male, but more narrowed posteriorly and with posterior margin more densely and longer setose medially; both sternite and tergite VIII (Figs 17, 18) conspicuously long and densely setose along posterior margin. Ovipositor (Fig. 19).

**Distribution.** Originally described from Australia (“Nová Hollandiát”), however occurring in Africa, and here very probably confined to the southern part of the Cape province of South Africa (see Remarks).

**Type material examined.** *Anthelephila cyanea* - Lectotype, female [herewith designated; mounted, together with the original triangular label, on rectangular label], labelled (Fig. 10a) “TYPE Hope Trans. Zool. Soc. I. P. 100. T.14 F.4 Coll. Hope Oxon. [both printed and handwritten, in fine red frame] ∨ Anthelephila cyanea Hope Z. J. [reddish label, handwritten, the last word illegible - see figure] ∨ TYPE Col : 1187 Anthelephila cyanea HOPE DEPT. OXFORD [both printed and handwritten, in black frame]” (OXUM). Paraplectotypes (lacking original labels): 2 females, mounted in the same way as lectotype (OXUM, BMNH); 1 female, mounted on triangular label, lacking head, with mouthparts mounted on separated label (OXUM).


**Remarks.** *Anthelephila cyanea* was described from Australia and never subsequently recorded until now. Having examined the type material, I found this species quite dissimilar to all its Australian congeners, placed in the here synonymized genus *Formicomus*, and identical in all characters with *F. caeruleus* (Thunberg, 1787) from southern Africa. Although there are examples of close relationships between the faunas of Australia and southern Africa (see Endrödi-Younga, 1978), *Anthelephila* does not seem to present this situation. Based on my studies, it appears to be rather derived genus, whose members colonized Australia from the Oriental region (see Distribution of the genus above). For these reasons, I believe that either the type locality of *A. cyanea* is erroneous, or the respective specimens were introduced to Australia.

Originally, *Formicomus caeruleus* was described by Thunberg (1787) in the genus *Notoxus* Geoffroy, 1762. Later, the species was listed under *Anthiscus* by Dejean (1837), and then transferred to *Formicomus* by Laferté-Sénéclaire (1849b, c). The examined syntypes are identical in all characters, including morphology of the prows of male sternite VIII (their apical portion is exposed in the male specimen) with the types and the specimens additionally examined of *Anthelephila cyanea*. Consequently, *F. caeruleus* is regarded as junior synonym of the latter species.

Considering the records of herein synonymized *F. caeruleus* from South Africa (Hille, 1950, 1961; Uhmann, 1981, 1983b, 1984; Chandler & Uhmann, 1984; Telnov 1998c), Namibia (Uhmann, 1985b, 1987b, 1995; Telnov 1998c), Botswana (Hille, 1986) and Angola (Hille, 1961), *A. cyanea* may appear like rather widespread species. However, I found the specimens originating from localities in the northern part of its range to be different from the typical *A. cyanea* in the more elongate elytra, regularly rounded, semicircular base of head, less enlarged distal antennomeres, moderately convex mesosternum (lacking paired impressions!) and in morphological details of the prows of sternite VIII (especially of their apical portion); the male sternite VIII of *F. caeruleus* figured by Hille (1950) shows similar differences. As solution of this problem was beyond the scope of the present paper, no attempt was made to continue with examination of additional specimens from different regions. Specimens of the questionable “northern form” were examined from the following localities (CKD, CUP, DSC): South Africa: Bloemfontein, Cape Town-Kloof Nek, Gross Barmen Resort, Kamieskron, Kuruman, Vioolsdrift, Winburg; Namibia: Noordoewer, Okahandja, Lüderitz, Warmbad, Windhoek.

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