Insemination and fertilization in the seed bug **Lygaeus simulans**
(Heteroptera: Lygaeidae)

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**Abstract.** In the laboratory, not all females of the seed bug *Lygaeus simulans* Deckert, 1985, produced fertilized eggs after copulation: 26.7% of the females were not inseminated and 5% were inseminated but did not lay fertilized eggs; only in 40% of the couples did copulation result in fertile eggs. The remaining 28.3% of couples refrained from mating. **Duration of copulation** was associated with insemination and fertilization: (i) fertile eggs were produced by only one couple that copulated for less than 60 min and all those that copulated for more than 360 min, (ii) probability of fertilization increased steadily with duration of copulation between 60 and 360 min, and (iii) duration of copulation was significantly different for couples that showed different insemination status.

A possible morphological explanation for this time dependency was revealed by examining the genitalia of 69 couples freeze-fixed in copula after different periods in copulation. Because of the intricate structure of the genitalia in *L. simulans*, a male takes a long time to maneuver its intromittent organ into the narrow insemination duct of the female. Only if completely inserted is the tip of the intromittent organ close enough for successful ejaculation of sperm into the spermatheca. The freeze-fixing experiment revealed that it usually took the male more than 30 min to locate the entrance to the insemination duct and another 30 min for full penetration. This explains why copulations that lasted less than 60 min failed, since insemination began only after intromission was complete.

The experiments, therefore, indicated that there is a relationship between the complex morphology of the genitalia and the low rates of insemination and fertilization in *L. simulans*.

**INTRODUCTION**

Females of many arthropod species have morphologically complex, tortuous genital systems that would appear to make insemination difficult (Eberhard, 1985, 1996). This may enable females to bias insemination rates by hampering the passage of the male’s intromittent organ to the spermatheca. In this way, females may prevent insemination by unsuitable males. Females may even exert control after insemination has occurred by preventing the fertilization of eggs with stored sperm (Eberhard, 1993; Rodriguez, 1994; Arnvist, 1997; Dickinson, 1997). Mechanisms excluding particular males as sires of the offspring have been discussed in the context of cryptic female choice (Eberhard, 1996) and male-female conflicts (Alexander et al., 1997).

In many species of lygaeid seed bugs, a male must completely insert its extremely long processus gonopori (intromittent organ) into the narrow and tortuous ductus receptaculi (insemination duct) of a female in order to reach the receptaculum seminis (spermatheca) and successfully deposit sperm (Ludwig, 1926: *Lygaeus* sp.; Bonhag & Wick, 1953: *Oncopeltus fasciatus*; Gschwentner, 1998: *L. simulans*). This study of *L. simulans* describes the mechanics of mating, records the duration of copulation associated with insemination and fertilization, and determines the influence of female genital morphology on the frequency of copulations that result in fertile eggs. The length of time required by a male to properly position its intromittent organ and inseminate a female was also determined.

**MATERIAL AND METHODS**

**Animals**

*L. simulans* is widespread throughout Eurasia, often occurring sympatrically with its sibling species *L. equestris* (Deckert, 1985; Lis, 1988; Gusev & Tatarnikov, 1991). Animals captured in a patch of steppe vegetation on Mount Hundsheim, Austria, were the progenitors of the culture established in the laboratory in the summer of 1995. Each generation takes about three months. The bugs were kept at a temperature of 28°C, a photoperiod of 16L : 8D and supplied with distilled water and peeled sunflower seeds. Only adult bugs that were between two weeks and two months old (the period of highest fecundity), which were reared individually from the last instar stage, were used in the experiments. Two to six days after they were laid, fertile eggs turn from white to red. This change in colour was used to distinguish fertile from infertile eggs. Fertilization success was recorded qualitatively (yes/no), not quantitatively, because either all or none of the eggs laid by a female were fertile. Quite a high percentage of the couples did not mate despite being sexually mature. In most cases, males tried to mate but were rejected by the females. Copulation attempts were not recorded.

When not in use, the male intromittent genitalia are contained within the conspicuous genital capsule at the end of the abdomen of males. The phallus consists of the proximal phallobase and the distal aedeagus, which bears the sclerotized processus gonopori of about 7 mm length. The ejaculatory duct winds through the phallus and opens at the tip of the processus. Prior to copulation, hemolymph pumped into the phallus causes it to unfold out of the capsule and inflates the aedeagus (cf. Ludwig,
After mates have assumed the typical tail-to-tail copulation posture, the phallus penetrates the female genital opening up to the phallobase until the aedeagus with the processus gonopori lies entirely inside the female bursa copulatrix. The narrow female ductus receptaculi originates in the bursa opening up to the phallobase until the aedeagus with the processus gonopori completely penetrates the ductus receptaculi. Only if thus inserted will the processus tip be sufficiently close to ejaculate sperm into the storage tube of the receptaculum seminis (Ludw, 1926: Figs 6, 8). After mates have assumed the typical tail-to-tail copulation posture, the phallus penetrates the female genital opening up to the phallobase until the aedeagus with the processus gonopori lies entirely inside the female bursa copulatrix.

Copulation experiment

The relation between the duration of copulation and the subsequent rates of insemination and fertilization was determined by allowing each of 120 couples, made up of mates taken at random from the stock of virgin animals, to copulate once in a Petri dish. Couples were observed continuously and duration of copulation recorded. When a couple ceased copulating or failed to mate within two hours, the male was removed from the dish. For the following three weeks, each female’s production of fertilized and unfertilized eggs was monitored, after which the females were fixed in 70% ethanol. Whole mounts of their receptacula seminis were embedded in Euparal and screened for the presence of sperm.

Freeze-fixing experiment

The relation between duration of copulation and the intromission depth of the processus gonopori inside the female ductus receptaculi was determined using couples in plastic mesh boxes (3 cm Ø, 0.5 cm high) allowed to copulate for a preset time: 60 couples each were allotted 30 min of copulation (a duration that previously had never resulted in fertile eggs – see also Fig. 2) and another 60 min (apparently the minimum duration for successful fertilization of eggs – see also Fig. 2). Half of the females in the 30 min set and half of those in the 60 min set were virgin; the other 30 in both sets had already been inseminated before the experiment. After the allocated periods had elapsed, plastic boxes with bugs still engaged in copula were swiftly placed in liquid nitrogen for a few seconds. Those that had not copulated after two hours had elapsed or had separated prematurely were discarded. Boxes with freeze-fixed couples were then placed into a mixture of 70% ethanol and 3% acetic acid at –18°C for two weeks. Whole mounts were later produced of the combined male-female genitalia and embedded in Euparal. Depth of penetration of the ductus receptaculi by the processus gonopori was recorded and classified.

RESULTS

Copulation experiment

Of the 120 virgin couples, 86 mated. They copulated for periods that ranged from 2 min to 16 h 51 min (mean duration 253.05 min, s = 242.09 min); most of the copulations lasted for short periods (Fig. 1). While it was always the male that approached the female and initiated copulation, it was not clear which sex terminated the mating. In general, mates remained motionless during long-lasting copulations and were restless during short copulations. Forty-eight females subsequently laid fertile eggs. The relation between fertilization and duration of copulation is given in Fig. 2: only one copulation that lasted for < 60 min and all copulations ≥ 360 min resulted in fertilization, while in those that lasted between 60 min and 360 min, the probability of fertilization (sperm present in the receptaculum seminis) increased, the longer the copulation lasted. The duration was significantly longer in those copulations that resulted in fertilization compared to those that did not (Kruskal-Wallis Test, χ² = 45.48, df = 1, p < 0.0001). A logistic regression model was fitted to the data and gave a better fit than the null model at the p < 0.0002 level (Chi-Square Test); probability of fertilization was negligible following copulations lasting less than 60 min and high for those that lasted more than 250 min (Fig. 3).

In 32 of the 86 mated females, there was no sperm in the receptaculum seminis. In a further 6 females, copulation resulted in insemination but not in fertilization. The remaining 48 females laid fertilized eggs. Therefore, lack of insemination can be due to either copulation fails to result in insemination (37% of the females) or insemination fails to result in fertilization (7% of the females).

Duration of copulation differed significantly between the three categories of couples (Kruskal-Wallis Test, χ² = 51.14, df = 2, p < 0.0001). Multiple comparisons showed that the duration of copulation for the 32 non-inseminated females was significantly different from that for the 48 fertile females (Mann-Whitney Test, two-tailed, p < 0.0001); in addition, that for the 32 non-inseminated females was significantly different from that for the 48 inseminated but infertile females (Mann-Whitney Test, two-tailed, p < 0.008). However, the duration of copulation for the 6 inseminated but infertile females was not significantly different from that for the 48 fertile females (Mann-Whitney Test, two-tailed, p > 0.0125). Since four simultaneous comparisons were made on the same data, the significance level was adjusted to a testwise error of α = 0.0125, according to the Bonferroni method (Sokal & Rohlf, 1995).
Fig. 2. Duration of copulation and fertilization in *Lygaeus simulans*. Duration was significantly longer in copulations that resulted in fertilization (Kruskal-Wallis Test, \( \chi^2 = 45.48, \text{df} = 1, p < 0.0001 \)).

**Freeze-fixing experiment**

Whole mount preparations of the male-female genitalia complexes of pairs in copula showed that the long ductus receptaculi is separated from the receptaculum seminis by a bell-shaped widening with a pore at its center (Fig. 6). This access pore into the receptaculum looks like a valve; however, it was not possible to determine whether sperm was stored only on one side of the valve. The receptaculum seminis consists of two sections: proximally, a short, translucent conduit of four to five tight spirals just after the access pore; distally, a conspicuously large, highly contorted, cuticularized sperm storage tube (Figs 4-7).

The anatomy of the spirals effectively and consistently prevents any direct contact between the processus gonopori and the sperm storage tube: the spirals are apparently too tightly-wound for the processus to maneuver through them; reversal of the direction of the coils after each spiral (Fig. 6) is a further obstruction. While the processus tip sometimes penetrated past the first and second spiral, it never extended past the third (Figs 4-7). Consequently, the processus ejaculates sperm into the ductus receptaculi (internal coupling) had not occurred. Even when the processus gonopori entered the ducus, the proximity of its tip to the receptaculum seminis at the time of freeze-fixing was highly variable (Figs 4–7). Five insertion depths (IDs) were distinguished:

- **ID 0**: the processus gonopori had not entered the ductus receptaculi in the allocated time. No whole mounts could be produced because ID 0 does not result in internal coupling of the genitalia: the processi slipped out of the bursae during dissection. That is, apart from intromission into the bursa copulatrix, copulation does not necessarily entail internal coupling.

- **ID 1**: the processus gonopori barely entered the ductus receptaculi (Fig. 4).
- **ID 2**: the processus tip reached midway into the ductus and was equidistant from the bursa and the access pore at the instant of freeze-fixing (Fig. 5).
- **ID 3**: the processus tip reached the end of the ductus and lay within the bell-shaped widening of the access pore (Fig. 6).
- **ID 4**: the processus tip had entered the access pore and reached the first, second or third turn of the spiral (Fig. 7).

The 30 min copulations with virgin females resulted in the largest number of ID 0 cases (non-intromission into the ductus receptaculi) and the smallest number of ID 4 cases (deepest penetration). The 60 min copulations with non-virgin females, in contrast, resulted in the smallest number of ID 0 cases and the largest number of ID 4 cases.
Figs 4–7: Lygaeus simulans. Depth of penetration into the female genitalia by the male intromittent organ; processus gonopori proximally cut off. 4 – processus gonopori barely inside ductus receptaculi (cf. ID 1); 5 – processus midway in ductus (cf. ID 2); 6 – processus tip at access pore (cf. ID 3); 7 – processus past access pore in spiral (cf. ID 4). For frequencies of IDs see Table 1. Abbreviations: a = access pore, d = ductus receptaculi, p = processus gonopori, s = spiral of the receptaculum seminis, t = sperm storage tube of the receptaculum seminis. Scale bars: 0.4 mm (4, 5, 7); 0.16 mm (6).

Table 1. Lygaeus simulans, five classes of intromission depth (ID 0 – ID 4) of the male intromittent organ within the female genitalia.

<table>
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<th>ID 0</th>
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<tr>
<td>virgin 30 min</td>
<td>9</td>
<td>4</td>
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<tr>
<td>virgin 60 min</td>
<td>4</td>
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<td>3</td>
<td>2</td>
<td>7</td>
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<tr>
<td>non-virgin 30 min</td>
<td>3</td>
<td>1</td>
<td>3</td>
<td>2</td>
<td>9</td>
</tr>
<tr>
<td>non-virgin 60 min</td>
<td>1</td>
<td>0</td>
<td>3</td>
<td>2</td>
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The relation between depth of penetration and duration of copulation (30 or 60 min) on the one hand and the virginity status of the females (virgin or non-virgin) on the other, was tested. For this test, only two classes of penetration were used, formed out of the original five ID classes: shallow penetration (ID 0/ID 1) and deep penetration (ID 3/ID 4). Since five simultaneous comparisons were made on the same data, the significance level was adjusted to a testwise error of α’ = 0.01 according to the Bonferroni method. Applying a “Pearson” chi-square test for goodness of fit to the data (Sokal & Rohlf, 1995) re-
vealed a non-random distribution of depth of penetrations both in relation with duration of copulation and the virginity status of females (χ² = 19.63, df = 4, p < 0.0006). Deep penetration was significantly more frequently observed when the duration of copulation was long and with non-virgin females. However, when the data were additionally subjected to four Fisher's exact tests of independence to investigate more closely the relation between penetration depth and duration of copulation and between penetration depth and virginity status, most differences were insignificant. The penetration depth (ID 0/ID 1 as opposed to ID 3/ID 4) recorded for the 30 min copulations with virgin females differed significantly from that recorded for the 30 min copulations with non-virgin females (two-tailed, p < 0.004). It was not significantly different in the 60 min copulations with virgin females compared with the 60 min copulations with non-virgin females (two-tailed, p > 0.01), or in the 30 min copulations with virgin females compared with the 60 min copulations with virgin females (two-tailed, p > 0.01), or in the 30 min copulations with non-virgin females compared with the 60 min copulations with non-virgin females (two-tailed, p > 0.01).

Whole mounts of deep/deepest penetration (ID 3/ID 4) often showed that at the instant of freeze-fixing, sperm strands were in the process of being ejaculated out of the processus gonopori tip into the sperm storage tube of the receptaculum seminis. A “Pearson” chi-square test for goodness of fit confirmed that ejaculation was significantly more frequent in the copulations of long duration and in matings involving non-virgin females (χ² = 21.1, df² = 3, p < 0.0001).

DISCUSSION

Lygaeid bugs remain in copula for long and variable periods of time (Ludwig, 1926; Andre, 1934; Hopp, 1988). In L. simulans, the longest copulation lasted for 1011 min and the mean duration was 253.05 min. Only 4% of the females that copulated for ≤ 60 min subsequently produced fertile eggs, whereas 100% of the females that copulated for ≥ 360 min did. In addition, the probability of a female being fertile increased steadily with the duration of copulation between 60 and 360 min (Figs 2–3).

Of the 86 females that mated, only 54 were inseminated. Six of the 54 females did not produce fertile eggs. As the ovaries of these females contained eggs when dissected, it is assumed they refrained from using the stored sperm for egg fertilization. It has been argued that a sperm storage organ and delayed fertilization enables females to accept or reject sperm following insemination and deprives males of direct influence on whether their sperm fertilizes eggs (Fowler, 1973; Gromko et al., 1984; Eberhard, 1996, 1997; Alexander et al., 1997; Brown et al., 1997; Dickinson, 1997). Remarkably, 22 out of the 86 couples copulated for several hours longer than the 6 h needed for successful fertilization (Fig. 1). According to the cryptic female choice theory, extended copulation may be a strategy used by males to induce females to utilize their sperm (Thornhill, 1983; Eberhard, 1985, 1996, 1997). Alternatively, long copulations may serve mate-guarding (Alcock, 1994). Although females were only exposed to one male, the mate-guarding hypothesis may still apply, since males may have sensed males in adjacent Petri dishes.

The duration of copulation was associated with whether the females were not inseminated, inseminated but not fertilized or fertilized. This supports the time-dependency of insemination and fertilization. The freeze-fixing experiment revealed that insemination is time-dependent as it takes most males at least 30 min to locate, with the tip of their processus gonopori (intromittent organ), the entrance to the female ductus receptaculius (insemination duct) inside the bursa copulatrix. Alternatively, searching for the ductus entrance may not be the problem and the male’s intromittent organ remains inside the bursa copulatrix for about 30 min before penetrating the ductus. After this, it takes approximately another 30 min for the processus to penetrate the ductus, pass the access pore and reach the tightly-wound spiral section of the receptaculum seminis (spermatheca). Only if inserted in this way does the tip of the processus lie close to the storage tube of the receptaculum seminis, into which the sperm is ejaculated (Figs 4–7, Table 1). Insemination is therefore unlikely to occur within the first hour of copulation in L. simulans. Similarly, a previous study on the sibling species L. equestris found that matings interrupted after 15, 30 and 60 min resulted in average fertilization rates of 2%, 17.6% and 45.9%, respectively. The long copulations (≥ 15 h) in this species are thought to indicate a male mate-guarding to forestall sperm displacement. When given the choice, the males preferred gravid to virgin females (Sillén-Tullberg, 1981). This is supported by the results from the freeze-fixing experiment: intromission was achieved faster in non-virgin females.

In L. simulans, the proper positioning of the male intromittent organ inside the female insemination duct is a prerequisite for successful sperm transfer (cf. Aedes aegypti: Gwadz et al., 1971; Gwadz, 1972). External coupling of the genitalia is achieved after the parameres of the male have seized the ovipositor, the phallus has penetrated the female genital opening and the aedeagus with the processus gonopori lies entirely inside the bursa copulatrix (Ludwig, 1926; Bonhag & Wick, 1953). However, the results of the freeze-fixing experiment clearly indicate that external coupling of the genitalia did not necessarily result in the processus gonopori entering the ductus receptaculi (frequency of ID 0; Table 1). If it does, the processus gonopori may only reach the ductus or the access pore separating the ductus from the receptaculum seminis. It is not clear whether incomplete intromission (e.g. ID 1, ID 2) is accompanied by insemination or whether sperm ejaculated into the ductus can eventually reach the sperm storage tube of the receptaculum. In the case of complete intromission and insemination, the tip of the processus reaches the spiral, which prevents it coming into direct contact with the sperm storage tube (frequencies of ID 1 – ID 4; Table 1). Functionally analogous to
the spiral in *L. simulans* is the “ampulla” structure that serves the same function in the chrysomelid beetle *Chelymorpha alternans* (Rodriguez, 1994; Rodriguez, Eberhard & Windsor, in prep., cited after Eberhard, 1996, pp. 351–353).

The morphologies of the ductus receptaculi and the receptaculum spiral exert a passive control over genital access in *L. simulans*. In addition, the longitudinal musculature enveloping the spiral and part of the bell-shaped widening that houses the access pore may actively control genital access (Gschwentner, 1998). Such passive and active control exist in various chrysomelid beetles: the spermathecal muscles actively block eversion of male genitalia (Dickinson, 1997) or insemination is passively prevented as the male fails to completely thread its long, thin intromittent organ up the tightly-coiled female spermathecal duct (Rodriguez, 1994).

The structure of the female genitalia (ductus, access pore, spiral) serves as an obstacle to the male’s intromittent organ up to the tightly-coiled female spermathecal duct (Rodriguez, 1994; Rodriguez, Eberhard & Bervache, 1996; Eberhard, 1997; Gschwentner, 1998). Such passive and active control over genital access (Gschwentner, 1998) may actively control eversion of male genitalia (Dickinson, 1997) or insemination is passively prevented as the male fails to completely thread its long, thin intromittent organ up the tightly-coiled female spermathecal duct (Rodriguez, 1994).

The structure of the female genitalia (ductus, access pore, spiral) serves as an obstacle to the male’s intromittent organ and thus probably biases insemination and fertilization rates. The long and winding insemination duct even continuing into a secondary tortuosity (spiral) before connecting to the remote receptaculum seminis may be a means of controlling the entry of sperm rather than insemination efficiency. Therefore, females of *L. simulans* obviously exert a mechanical cryptic female choice via the morphology of their genitalia.

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